

Medical Science Sample

Case Report

Acute Appendicitis Masquerading as Distal Intestinal Obstruction Syndrome in Adult

Cystic Fibrosis

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Overshadowed by ~~S~~ino-pulmonary infections, ~~c~~Cystic ~~f~~Fibrosis (CF) commonly affects gastrointestinal organs because of secretory and motility dysfunction. Infrequently, the ~~resulting~~ changes ~~can result~~ ~~in~~cause ~~d~~istal ~~i~~ntestinal ~~o~~bststruction ~~s~~ndrome (DIOS), an ~~more and more~~increasingly diagnosed gastrointestinal ~~condition~~entity in adult ~~Cystic Fibrosis~~CF patients. We present ~~thea~~ case ~~of a~~ 22-year-old ~~man~~le who presented to our hospital with right lower quadrant abdominal pain. ~~with~~~~Despite the~~ suspicion of acute appendicitis, ~~the patient and~~was subsequently diagnosed ~~as with~~ DIOS. Our case highlights the importance of ~~considering~~ DIOS as ~~a~~ differential diagnosis ~~of for~~ right lower quadrant abdominal pain in CF patients, especially ~~for by~~ physicians working at community hospitals ~~that~~which may not have a ~~C~~Fyctic ~~F~~ibrosis care program available.

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1. Introduction

Cystic ~~F~~ibrosis (CF) is a genetic disease ~~of that affects~~ multiple organs. ~~With~~~~Because of~~ ~~advancements~~ing in the ~~management~~ing of CF ~~patients~~, patients ~~can~~ now ~~often survive~~ ~~become to~~ adulthood [1]. ~~However, the~~ ~~improved~~ life expectancy among adult CF patients has ~~given rise~~ed to ~~an increase in~~ extrapulmonary, notably gastrointestinal, ~~manifestations~~, which ~~did not happen~~ ~~was~~ previously ~~uncommon~~. Distal ~~i~~ntestinal ~~o~~bststruction ~~s~~ndrome (DIOS) continues to be a rising complication in adult CF patients, presenting ~~as with~~ acute abdominal pain ~~like and~~ ~~mimicking~~ an acute abdominal emergency.

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2. Case Report

A 22-year-old Turkish-origin ~~man~~le with a past medical history of ~~Cystic Fibrosis~~CF presented with a one-day history of right lower quadrant abdominal pain. He described ~~a~~ sharp periumbilical pain that continued to worsen, which then shifted to ~~the~~ right lower quadrant ~~of the~~ abdomen. Prior to the onset of the abdominal pain, he reported ~~experiencing~~ nausea and anorexia for three days. His last bowel movement was two

days prior to admission. Upon reviewing the patient's past history, it was noted that he had several episodes of pneumo-nia, for which he was appropriately treated with antibiotics. Notably, no history of constipation or recurrent abdominal discomfort was reported prior to this. At home, the patient was prescribed Albuterol inhalation as needed, Dornase Alfa inhalation, Aztreonam lysine nebulization, 500 mg Azithromycin three times a week, Lansoprazole, Lumacaftor-ivacaftor twice a day, Lipase-protease-amylase capsule three times a day, and a multivitamin capsule once a day. The patient was also diagnosed with Cystic Fibrosis CF at the age of four, and this disease progressed to exocrine pancreatic insufficiency, which was being treated with pancreatic enzymes. On abdominal examination, he was found to have had diminished bowel sounds and tenderness on right lower quadrant with equivocal rebound tenderness on the right lower quadrant. Laboratory analysis showed leukocytosis (white blood cell count, WBC 13.0 mm/K3; Neutrophils count, 62%) with a normal differential. He had no electrolyte imbalances. Computed Tomography (CT) of the Abdomen revealed thickening, and edema around the terminal ileum, inflammatory changes in the a-colon with inflammatory changes, free fluid in the right paracolic gutter adjacent to the cecum, an appendix measuring 5.3x4.6 mm, and reactive lymph nodes (Figures 1 and 2).

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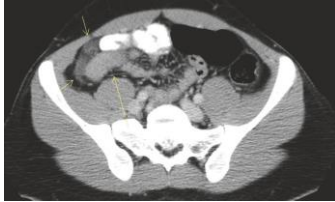


FIGURE 1: Axial abdominal computed tomography scan depicting thickening around the terminal ileum and colon (yellow arrows) along with extraluminal fluid and reactive lymph nodes.

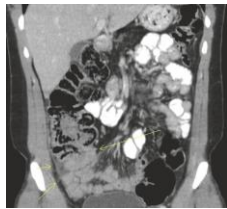


FIGURE 2: Coronal view computed tomography scan with showing thickening of the ileum with a distended appendix (yellow arrows).

measuring 5.34.6 mm, and reactive lymph nodes (Figures 1 and 2). Due to extraluminal fluid and cecal wall edema with inflammation, early acute appendicitis could not be excluded as a possible diagnosis. Surgical intervention was performed, which revealed a ruptured microperforation of a cecal diverticulum and a distended appendix in chronic adhesions, for which he required an appendectomy and partial cecectomy with an intact ileocecal valve (IC valve) valve. Postoperatively, he was diagnosed with DIOS and was subsequently started on pPolyethylene glycol. The patient made an unremarkable recovery and was discharged. He was followed up in the

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outpatient clinic ~~without and did not have any~~ recurrence of any symptoms.

3. Discussion

~~Distal Intestinal Obstruction Syndrome (DIOS) was previously called known as~~ Meconium Ileus ~~-equivalent in the past, described is characterized~~ by the collection of viscid fecal material within the lumen combined with sticky mucoid intestinal content adherent to the intestinal wall of the terminal ileum and cecum [1]. Perez-Aguilar et al. reported ~~that thea~~ prevalence of ~~DIOS was~~ 19.5% (mean age 20.6 years) among 46 CF patients in a retrospective analysis, while Dray et al. ~~conducted a cross-sectional study-reporting~~ a 15.8% (mean age 28.9 years) prevalence ~~in among~~ 171 CF patients ~~in a cross-sectional study~~ [2, 3]. ~~Despite the~~ ~~Though there continues to be a~~ limited assessment ~~of-on~~ the prevalence of DIOS in adult CF ~~patients~~, DIOS is ~~consideredmore~~ common among adults ~~compared to than among~~ children ~~due to because of increased~~ disease progression.

Defective intestinal chloride and water secretions into the gut, luminal acidity, and loss of bile salt all contribute to the

development of DIOS [1]. These patients characteristically present with right lower quadrant pain, nausea, abdominal distension, and failure to pass stools or flatus [1, 3]. In some patients, a palpable right lower quadrant mass ~~can may be appreciated present~~ that may be confirmed on abdominal ~~radiography X-ray~~ [1]. Though abdominal ~~X-rays are radiography is~~ recommended to aid in the diagnosis of DIOS, ~~they are it is~~ inadequate in differentiating ileus from other causes of abdominal pathologies that may present in ~~Cystic Fibrosis CF~~ patients [4]. Due to ~~the proximity of the anatomical locations proximity~~, as well as the overlapping clinical presentations, appendicitis and intussusception may mimic DIOS, which further leads to diagnostic uncertainty. ~~Overlap of several intra-abdominal pathologies in CF increases the risk of misdiagnosis, especially with for acute appendicitis,~~ as these ~~patient's~~ underlying pathologies may be masked ~~in patients~~ with pulmonary infections² ~~using~~ antibiotics [5, 6].

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Osmotic laxatives are the cornerstone of bowel regimens for the treatment of DIOS. The most commonly prescribed ~~laxative is p~~Polyethylene Glycol, (PEG)-~~administered~~ at a dose of 20–40 ml/~~kKg/hH~~, ~~up twith~~ a maximum of 1 ~~lL/kg/h~~ for a total of 8 hours, ~~resulting in a achieving~~ fecal effluent consisting of clear fluid, along with ~~the~~ resolution of abdominal pain and constipation [1, 6]. If the diagnosis remains unclear, and thus, requires surgical intervention, ~~IC ileocecal~~ valve resection should be considered to prevent the ~~development and~~ recurrence of intestinal obstruction sequelae ~~and growth~~, especially in adolescents [7].

With the increase in immigration of foreigners ~~into through~~ America, inner-city and community hospitals may not be ~~sufficiently~~ equipped with a ~~Cystic Fibrosis CF~~ care center; ~~moreover, nor may~~ these hospitals ~~may not~~ have programs in provision, with expertise available to other clinicians involved in patient care.

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Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors' Contributions

All authors contributed to the revision and approval of the manuscript.

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Life Sciences Sample

Case Report

Methylmalonic Acidemia with Novel *MUT* Gene Mutations

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A 5-year-old boy presented with recurrent episodes of fever, feeding problems, and lethargy, ~~from since~~ the age of 11 months, and poor weight gain. He was admitted to our hospital and evaluated for metabolic disorders; subsequently, he ~~causes and was~~ diagnosed with methylmalonic acidemia (MMA). He was treated with vitamin B12 and carnitine supplements and has been followed-up for the last 3 years. Mutation analysis by next generation sequencing (NGS), supplemented with Sanger sequencing, revealed two novel variants in exon 5 and exon 3 of the *MUT* gene responsible for the methylmalonic acidemia MMA in exon 5 and exon 3. Recently, he had developed dystonic movements including orofacial dyskinesia. With the advent of NGS, judicious use of NGS with Sanger sequencing can help identify causative and possibly pathogenic mutations.

1. Case Presentation

A 5-year-old child boy presented for the first time at the age of 11 months, with complaints of fever, vomiting, poor feeding, and lethargy for the first time at the age of 11 months. We observed that the patient had pallor and tachypnea and was drowsy. Further evaluation was suggestive of high anion-gap metabolic acidosis with ketonuria (urine ketones 3+) and with normal electrolytes, blood sugar (94 mg/dL), vitamin B12, and homocysteine levels. Plasma ammonia and plasma lactate were 118 units, and plasma lactate was 2.9 units, respectively. Transcranial magnetic stimulation TMS results were normal, but gas chromatography mass spectrometry analysis of urine GCMS revealed elevated 3-OH propionic acid [12.39 retention time (RT)] as well as elevated methyl malonic acid levels [16.92 RT, Suppl Figure 1, in Supplementary Material available online at <https://doi.org/10.1155/2017/8984951>]. Since then, the child patient was on a low-protein diet, and carnitine, biotin, thiamine, and vitamin B12 injections. The child was thereafter admitted to the hospital on seven multiple occasions (7 times) with acute decompensation and managed as per protocol. Mutational analysis was sent for methylmalonic acidemia (MMA) which showed a single heterozygous missense variant c.976 A>G (p.Arg326Gly) in exon 5 of the *MUT* gene (genomic coordinates: chr 6: 49421405); as a variant of uncertain significance. Chromosomal microarray analysis ~~done~~ did not reveal any major deletion or duplication that which could disrupt the gene. Since exon 3 and exon 6 were not adequately covered by next generation sequencing (NGS), further evaluation by Sanger sequencing for targeted exons was performed, and a second mutation in exon 3 c.753 G>A (p.=) was identified. The variants were predicted as found to be damaging by the SIFT database score (Suppl data) and as they were also predicted to be deleterious by Polyphen-2 and Mutation-Taster, but

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they were ~~and absent not found~~ in the ExAC database. ~~Brain magnetic resonance image MRI brain of the patient (done at from~~ the age of ~~four~~4 years) ~~was showing~~ multifocal cystic encephalomalacic changes with surrounding gliosis in deep white matter predominantly in frontoparietal regions (Figure 1). ~~During in the latest admission of the patient to the hospital, we observed child was found to have~~ fresh neurological findings in the form of perioral tremors, generaliz~~ed~~ed hypertonia, and generaliz~~ed~~ed dystonia with clonus with exaggerated deep tendon reflexes. ~~The patient He~~ was treated with intravenous dextrose and sodium bicarbonate and was continued on carnitine and ~~injection of~~ vitamin B12 ~~injections~~. Plasma ammonia ~~and plasma lactate were was~~ 18 units and ~~lactate level was~~ 4.9 units, ~~respectively~~. ~~Brain magnetic resonance image MRI brain of the patient was repeated and~~ revealed bilateral basal ganglia hyperintensities, suggestive of metabolic stroke. After the subsidence of acute crisis, he was discharged on carnitine, ~~injection of~~ vitamin B12, ~~injections~~, and trihexyphenidyl. ~~His p~~Parents were counseled regarding ~~the~~ prognosis and for prenatal diagnosis ~~for next subsequent pregnancies~~.

2. Discussion

MMA presents with lethargy, acidosis, hypoglycemia/ hyperglycemia, ketosis, and recurrent episodes. ~~MMA due to MUT gene mutations usually leads to severe phenotypes due to MUT gene mutations~~, and around 35–40% of cases are due to ~~novel~~ mutations [1, 2]. ~~There can be Missense or nonsense mutations, deletions, insertions, and so on in the MUT gene and so on can leading to a clinical phenotype.~~



Figure 1: ~~The Brain magnetic resonance image MRI brain of in~~ the -child -with MUT-related ~~methyilmalonic acidemiaMMA~~ showing ~~predominant frontoparietal abnormalities in the~~ form of encephalomalacia and gliosis.

The advent of NGS technology has enabled better characterization of mutations in several populations. However, Sanger sequencing remains ~~a~~ useful adjunct in molecular testing ~~in of~~ these cases. ~~It is required to find mutations when there is a strong clinical suspicion for them. Sometimes in NGS, due to because of~~ incomplete coverage of the exons ~~by NGS, Sanger sequencing is required to find mutations, if there is strong clinical suspicion. In this study, by using both the techniques. By careful use of both techniques, we could found~~ the two *MUT*

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variants responsible for- MMA in the patient~~the clinical condition~~. In a Saudi study on 60 patients of MMA patients, nonsense, missense, and frameshift mutations were detected across the *MUT* gene [3]. Another study in 43 Chinese patients identified 8 recurrent mutations and 10 novel mutations [4]. A previous Indian study in 15 patients with of clinically diagnosed MMA identified one novel exon 12 mutation in the *MUT* gene with predicted pathogenicity. In this caseHere, we identified two novel variants, one in exon 3 and another in exon 5 of the *MUT* gene. Both were labelled as variants of unknown significance (*VUS*). The exon 3 variant is a synonymous variant, and a different nucleotide change c.753 G>C (p.Lys251Asn) has been reported earlier in ClinVar. Some synonymous variants can also affect the splicing or protein function and lead to clinical phenotypes. The identified exon 5 variant is novelnew, but another close variant c.977 G>A (p.Arg326Lys) has been reported in ClinVar. The variants were found to be deleterious on bioinformatic analysis and were absent not found in the ExAC database. Both variants identified in the present case could possibly explainbe responsible for the phenotype of MMA phenotype in the child. *MUT*-related MMA has poor prognosis in most cases. Specialized diet and supplements may not improve the outcomes, even if MMA is diagnosed early. Early recognition and appropriate treatment of acute crises are necessary. Metabolic stroke can sometimes occur in the absence of acute metabolic decompensation, so meticulous neurological examination at every each visit is useful. The treatment options for therapy include early liver transplantation [5] and possibly gene therapy in the future. Genetic counseling and prenatal diagnosis could help these families of the patients in making reproductive decisions.

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Conflicts of Interest

The authors declare that they have no conflicts of interest.

Acknowledgments

The authors would like to acknowledge Dhiti Omics Technologies Pvt Ltd for help in mutation analysis.

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Physical Science Sample

Structural Prediction prediction of Bisbis(di-p-anisole)-1,4-azabutadiene-bis(triphenylphosphine)ruthenium(II) Using ³¹P NMR Spectroscopy

Author Details

Abstract¹

The present paper reports the use of ³¹P NMR spectroscopy to predict the isomers structures of [bis-4-methoxy-phenyl-[3-(4-methoxy-phenyl)-allylidene]-amino]-bis(triphenylphosphine)ruthenium(II), also known as bis(di-p-anisole)-1,4-azabutadiene-bis(triphenylphosphine)ruthenium(II), complexes. The complexation reaction was carried out using (di-p-anisole)-1,4-azabutadiene (compound 1), triphenylphosphine, and ruthenium chloride in 2:2:1 ratio under refluxing conditions of (di-p-anisole)-1,4-azabutadiene (compound 1), triphenylphosphine (PPh₃), and ruthenium chloride in the ratio of 2:2:1 for five 5 hours. The formation of the In addition, ruthenium(II) complexes were was further confirmed by also characterized using FTIR and UV-Vis spectroscopy analyse to support the formation of ruthenium(II) complexes. The results of ³¹P NMR spectroscopy study on ruthenium(II) complexes suggested indicated the presence of that there are three isomers present after the complexation reaction.

Keywords:

¹ NMR, nuclear magnetic resonance;

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1. Introduction

Nuclear magnetic resonance (NMR) spectroscopy is an essential instrument-analytical tool in the field of chemistry as it can help determine-elucidate the structure of a molecule, identify-detect the presence of impurities in a sample, and determine the rates-of formation and-as-well-as-degradation of a compound. Even in 1970s, NMR has been used as early as in the 1970s already been used to determine-detect the cancer formation which had been identified to be offered a simple, fast, and low-cost method for this purpose identify-cancer-formation [1-3].

In As part of our long-term research interest-on the synthesis of in-ruthenium(II) complexes-synthesis, we used the (di-*p*-anisole)-1,4-azabutadiene (**1**) and triphenylphosphine (PPh₃) as the ligands to-for reaction react with ruthenium trichloride under reflux conditions. The resulting pProducts were formed, were checked-analyzed by using ³¹P NMR spectroscopy, and the spectral observations-results found-in the spectra are worth-to-be-discussed in the present communication.

For Inorganic inorganic chemists commonly use, using-of ³¹P NMR spectroscopy to identify the structure of a complex containing phosphine ligands is very common [4, 5]. ThFor example, this technique has been used well-known examples-is the use of ³¹P NMR spectroscopy to determine-elucidate the mechanism of Wilkinson hydrogenation mechanism-based on by identifying the coupling patterns among the phosphine ligands as well as those-and-also-the-coupling-constants-between the phosphine ligands as-well-as-and the rhodium(I) metal center [6].

2. Methodology

The ruthenium complexes were characterized using UV-Vis, FTIR, and ³¹P NMR spectroscopy. The IR spectra were recorded using-on a Thermo Scientific Nicolet iS10 spectrophotometer in-using KBr disc. The ¹H NMR spectrum for-of compound **1** and ³¹P NMR spectrum for-of the ruthenium(II) complexes were recorded using-on a JEOL JNM-ECA 500 spectrometer with TMS as an-the internal standard. The absorption spectra was-were recorded with-on a Jasco V-630 UV-Vis spectrophotometer.

2.1. To prepare Preparation of (4-Methoxy-phenyl)-[3-(4-methoxy-phenyl)-allylidene]-amine or (di-*p*-Anisole)-1,4-azabutadiene (**1**).

4-Methoxycinnamaldehyde-methoxycinnamaldehyde (1.62 g, 10.00 mmol) was dissolved in 10 mL of ethanol, and followed by the addition of 4-methoxyaniline (1.23 g, 10.00 mmol) which was then added to solution. The rReaction mixture was stirred and-to obtain a resulted-in-green-yellow solid, which. The solid was filtered, washed with 5 mL of ethanol, and dried *in vacuo*. The solid was purified by dissolving it in DCM and layered with hexane via slow diffusion. Yield: 2.368 g (88.7%); IR (KBr, cm⁻¹): 3036 (C-H stretching), 1627 (C=N- stretching), 1601 (C=C stretching, aliphatic), 1575 and 1468 (C=C stretching, aromatic), and 1110 (OCH₃ stretching); ¹H NMR (500 MHz, CDCl₃): δ: 8.25 (d, 1H, Hz, -CH=N-), 7.47 (d, 2H, Hz-), 7.18 (d, 2H, Hz-), 7.05 (t, 1H, Hz, H-C_α), 6.99 (m, 1H, H-C_β), 6.90 (d, 4H, Hz-), 3.83 (s, 3H, OCH₃), and 3.81 (s, 3H, OCH₃); UV-Vis (DCM, /nm): 273, 373; Anal. Calc. for C₁₇H₁₇O₂N (%): C, 76.38; H, 6.41; N, 5.24; Found (%): C, 76.75; H, 6.31; N, 5.05.

To prepare 2.2. Preparation of [Bis(4-methoxy-phenyl)-[3-(4-methoxy-phenyl)-allylidene]-amino]-bis-[triphenylphosphine]ruthenium(II) or Bis(di-*p*-anisole)-1,4-azabutadiene]-bis[triphenylphosphine]ruthenium(II) Complex-complex, es

RuCl₃·xH₂O (2.070 g, 1.0 mmol) and PPh₃ (0.525 g, 2.0 mmol) were added to a round-bottom flask containing 10 mL ethanol, and the mixture was then-refluxed. Compound **1** (0.316 g, 2.0 mmol) was then added to the round-bottom flask, and the mixture was refluxed again. The resulting pPale-maroon solids were-was-formed, filtered and washed with hexane, and the p-Precipitate was dried *in vacuo*: IR (KBr, cm⁻¹): 3034 (C-H stretching), 1661 (C=N), 1576 (-merged IR band of-for aliphatic and aromatic C=C stretching-from aliphatic and aromatic), 1469 (C=C stretching of aromatic ring), and 654 (Ru-C), and 577 (Ru-N); ³¹P NMR (202.5 MHz, CDCl₃): δ: 49.7 (d, 1P, Hz), 47.4 (d, 1P, Hz), 41.7 (d, 1P, Hz), 39.7 (d, 1P, Hz), 35.1 (s, Ph₃P=O), and 29.9 (s, 1P); UV-Vis (DCM) (λ): 321 and 382.

3. Results and Discussion

The Characterization of the ruthenium complexes were characterized was done using by UV-Vis, FTIR,

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and ^{31}P NMR spectroscopy. The IR spectra were recorded found on a by Thermo Scientific Nicolet iS10 spectrophotometer in using KBr discs. ^1H NMR spectrum for of compound 1 and ^{31}P NMR spectrum spectra for the ruthenium(II) complexes were recorded on a obtained through JEOL JNM-ECA 500 spectrometer with TMS as an internal standard. The absorption spectra were recorded with on a Jasco V-630 UV-Vis spectrophotometer.

The ^{31}P NMR spectrum of the ruthenium complexes (Fig. 1) shows appearance of two pairs of doublets and one singlet, indicating in the ^{31}P NMR spectrum for ruthenium complexes (Figure 1) indicate the presence of that there are three isomers (1:1:1 ratio) present induring the complexation reaction with the ratio of 1:1:1.

FigureFig. 1: ^{31}P NMR spectrum for of ruthenium(II) complexes.

The singlet at 29.88 ppm reveals that the two PPh_3 units are magnetically equivalent in the ruthenium(II) complex. The In this case, the two PPh_3 units are either located at the axial position and are, which is trans to each other (FigureFig. 2(a)) [7], or located at in the equatorial plane, which is only trans only to either one of the C atoms from in the $\text{C}=\text{C}$ bond or the N atom from in the $\text{N}=\text{C}$ bond (FigureFig. 2(b)).

FigureFig. 2: Postulated structures of (a) *trans*- and ((b) and (c)) *cis*-[bis(di-*p*-anisole)-1,4-azabutadiene]-bis[triphenylphosphine]ruthenium(II).

Meanwhile, a The pair of doublets at 41.84 and 39.74 ppm with a -coupling constant of 21 Hz is assigned to a the *cis*-isomer of the ruthenium(II) complex, as shown in FigureFig. 3(a). Lastly A, another pair of doublets at 49.80 and 47.36 ppm with a coupling constant of 38 Hz is assigned to a the *trans*-ruthenium(II) complex (FigureFig. 3(b)). The difference in-coupling between the ruthenium(II) complexes in FiguresFig. 3(a) and 3(b) is due to the positions of the PPh_3 ligands. The smaller coupling constant of τ , namely, 21 Hz is, was assigned to the *cis*-isomer because both the PPh_3 ligands are in the equatorial plane. Fig. 3(a) shows The presence of doublets, which are for assignable to the PPh_3 ligands in the complex is shown in Figure 3(a) because both the PPh_3 ligands are trans to different atoms, that is, (nitrogen and carbon) atoms. For In the ruthenium(II) complex (as shown in FigureFig. 3(b)), the two PPh_3 ligands are located at the axial position and are trans to each other. The Lastly, the single peak observed at 35.14 ppm is attributed to the presence of the triphenylphosphine oxide [8].

FigureFig. 3: Postulated structures of (a) *cis*- and (b) *trans*-[bis(di-*p*-anisole)-1,4-azabutadiene]-bis[triphenylphosphine]ruthenium(II).

On the other hand, theThe binding of compound 1 to the ruthenium(II) metal eentrecenter is can be confirmed using FTIR and UV-Vis spectroscopy. Comparison ofing the IR spectra between of compound 1 and the ruthenium complexes (FigureFig. 4) reveals that, the vibrations of $\text{C}=\text{N}$ and $\text{C}=\text{C}$ stretching bands bands are have been shifted after binding to the ruthenium(II) metal eentrecenter. The For $\text{C}=\text{N}$ stretching band, it shifted from 1627 cm^{-1} in the spectrum of compound 1 to 1661 cm^{-1} in the spectrum of the ruthenium complex [9, 10]. In contrast, whereas the for $\text{C}=\text{C}$ stretching, the IR band appears at 1601 cm^{-1} in the spectrum of compound 1 but it is not clearly shown detected in the spectrum of the complex because the IR bands of aliphatic and aromatic $\text{C}=\text{C}$ bands for aliphatic and aromatic were merging into one a single broad IR-band eentrecentered at 1576 cm^{-1} . Nevertheless, the two additional IR-peaks are present at 577 and 654 cm^{-1} in the finger-print region of the spectrum at 577 and 654 cm^{-1} indicating confirm the formation of the respective Ru-N and Ru-C bonds [11].

FigureFig. 4: IR spectra of (a) compound 1 (a) and (b) ruthenium(II)- complexes (b).

The complexation of compound 1 to the ruthenium(II)- metal eentrecenter is can be further supported by the UV-Vis data spectra as shown in FigureFig. 5. For In the case of compound 1, two absorption bands were are observed at 273 and 372 nm, which are assigned to the transition of the benzene ring and -transition of thej-imine group [12], respectively. After the complexation, both absorption bands show significant shifts to 321 and 382 nm, respectively, demonstrating the Significant shifts of these two absorption bands have proven compound 1 was successfully bound-binding of 1 to the ruthenium(II) metal eentrecenter via the nitrogen atom from in the $\text{C}=\text{N}$ group and the carbon atom from in the aliphatic $\text{C}=\text{C}$ aliphatic group in of the $\text{C}=\text{C}-\text{C}=\text{N}$ moiety.

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Figure 5: UV-Vis spectra of (a) compound 1 and (b) ruthenium (II) complex.

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4. Conclusion

Based on ^{31}P NMR spectral evidence, we confirmed from ^{31}P NMR spectrum has shown the presence of three isomers of the bis(di-*p*-anisole)-1,4-azabutadiene-bis(triphenylphosphine)ruthenium(II) complex, in the 1:1:1 ratio of 1:1:1. In addition, the data from IR and UV-Vis spectral data revealed the successful binding of compound 1 has bound to the ruthenium(II) metal center.

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